AN EXTREME LEARNING MACHINE CLASSIFIER FOR PREDICTION OF RELATIVE SOLVENT ACCESSIBILITY IN PROTEINS

Saras Saraswathi, Andrzej Kloczkowski and Robert L. Jernigan
Department of Biochemistry, Biophysics and Molecular Biology
L. H. Baker Center for Bioinformatics and Biological Statistics, Iowa State University
112 Office and Laboratory Building, Ames, IA, 50011, U.S.A.

Keywords: Relative solvent accessibility, Support vector machine, Neural network, Extreme learning machine, Prediction.

Abstract: A neural network based method called Sparse-Extreme Learning Machine (S-ELM) is used for prediction of Relative Solvent Accessibility (RSA) in proteins. We have shown that multiple-fold gains in speed of processing by S-ELM compared to using SVM for classification, while accuracy efficiencies are comparable to literature. The study indicates that using S-ELM would give a distinct advantage in terms of processing speed and performance for RSA prediction.

1 INTRODUCTION

Proteins perform a variety of important biological functions that are imperative to the wellbeing of all living things. Various factors determine protein functions, such as, its native structure, the information coded in its constituent amino acid sequences, its reactions to the surrounding solvent environment and the Relative Solvent Accessibility (RSA) values of its residues and. Evaluating RSA values will help to gain an insight into the structure and function of a protein.

Protein structures and other related values such as RSA can be experimentally determined by using NMR spectroscopy or X-Ray crystallography. But these methods can be expensive in terms of cost, time and other factors. There is an urgent need to process large amounts of data (spawned by advances in biotechnology) accurately and speedily in order to decipher the information buried in biological data, since it is impractical to do it manually. Computational methods such as machine learning algorithms provide an alternate way by which we can study this data in a cost and time efficient manner. Still, accuracies and processing efficiencies in existing methods are inadequate and there is a need for improvement. This study endeavours to attain a large gain in processing efficiencies.

RSA prediction has contributed to the study of protein functions in many applications; to determine protein hydration properties (Ooi, Oobatake, Namethy, & Scheraga, 1987), identify temperature sensitive residues that can be targeted for mutagenesis and to study contact residue information (Shen and Vihinen 2003), improve secondary structure prediction (Adamczak, Porollo & Meller, 2005) and for fold recognition and protein domain (DOMpro) prediction (Cheng and Baldi, 2006). RSA values can be used to gauge degree of solvent exposure of segments of globular proteins (Carugo, 2003), to find residues with potential structural or functional (ConSeq) importance (Berezin, Glaser, Rosenberg, Paz, Pupko, Fariselli, Casadio, & Ben-Tal, 2004), help with rationale design of antibodies and other proteins to improve binding affinities (David, Asprer, Ibana, Concepcion & Padlan, 2007). In general RSA values can help to achieve cost and time efficiencies in drug discovery processes and help to gain a better understanding of biological processes.

Probability profiles are used by Gianese, Bossa & Pascarella (2003) to predict RSA values from single sequence and Multiple Sequence Alignment (MSA) data. Singh, Gromiha, Sarai & Ahmad (2006) estimate RSA values from an atomic perspective. Pollastri, Martin, Mooney & Vullo

Saraswathi S., Jernigan R. and Kloczkowski A..
AN EXTREME LEARNING MACHINE CLASSIFIER FOR PREDICTION OF RELATIVE SOLVENT ACCESSIBILITY IN PROTEINS .
DOI: 10.5220/0003086803640369
In *Proceedings of the International Conference on Fuzzy Computation and 2nd International Conference on Neural Computation* (ICNC-2010), pages 364-369
ISBN: 978-989-8425-32-4

(2007) use homologous structural information to improve RSA prediction. In addition, tertiary structure predictions are increasingly being augmented and improved with information derived from secondary structure and RSA predictions. Zarei, Arab & Sadeghi (2007) find that pairs of residues can influence RSA prediction accuracy.

Knowledge-based tools which use machine learning techniques and statistical theory can be valuable in predicting RSA, especially in the absence of evolutionary information or where sequences are not well preserved. A number of computational methods have been used for RSA prediction, such as Neural Networks (NN) (Shandar and Gromiha, 2002; Adamczak et al., 2005; Cheng, Sweredoski, & Baldi, 2006; Huang, Zhu & Siew, 2006;). Pollastri, Baldi, Fariselli & Casadio (2002) use RSA values of residues for scoring remote homology searches and modelling protein folding and structure using a bidirectional recurrent neural network (ACCpro). Other methods include Information Theory (Manesh, Sadeghi, Arab & Movahedi, 2001), Multiple Linear Regression Methods (Pollastri et al. 2002; Wagner et al. 2005), Support Vector Machines (SVM) (Nguyen and Rajapakse 2005) and fuzzy *k*-nearest neighbour algorithm (Sim, Kim & Lee, 2005). Kim and Park (2004) have used the SVMpsi and long range interactions to improve RSA accuracy. Chen, Zhou, Hu & Yoo (2004) compare five different methods, decision tree (DT), Support Vector Machine (SVM), Bayesian Statistics (BS), Neural Network (NN) and Multiple Linear Regression (MLR) on the same data set in order to compare the capabilities of different methods in predicting RSA. They conclude that NN and SVM are among the best methods for RSA prediction.

More recently, Bondugula and Xu (2008) combine sequence and structural information to estimate RSA values (MUPRED) in order to predict RSA. Petersen, Petersen, Andersen, Nielsen and Lundegaard (2009) argue for the need of a reliability score (Z-score) for measuring the degree of trust that can be related to individual predictions. Meshkin and Ghafuri (2010) use a two-step approach, using feature selection on physico-chemical properties of residues and Support Vector Regression (SVR) to predict RSA.

We propose to use a new fairly new method called Sparse Extreme Learning Machine (S-ELM), based on neural networks, which is capable of extreme speeds compared to traditional neural networks while maintaining current classification accuracies.

This paper is organized as follows. Section 2 briefly discusses the S-ELM algorithm and characteristics of the RSA data. Section 3 discusses the results of this study with performance comparisons with SVM and NETASA methods followed by conclusions in Section 4.

2 METHODS AND DATA

2.1 Extreme Learning Machine

Single Layer Feed-forward Network (SLFN), with a hidden layer and an activation function possess an inherent structure suitable for mapping complex characteristics, learning and optimization. They have applications in bioinformatics for solving various problems like pattern classification and recognition, structure prediction and data mining. The free parameters of the network are learned from given training samples using gradient descent algorithms that are relatively slow and have many issues in error convergence. A modified SLFN model called an Extreme Learning Machine (ELM) has emerged recently (Huang, Zhu, & Siew 2006), where it has been proved theoretically that ELM can provide good generalization performance and overcome some of the problems associated with traditional NNs such as stopping criterion, learning rate, number of epochs and local minima. ELM has good generalization capabilities and capacity to learn extremely fast. The input weights are chosen randomly but the output weights are calculated analytically using a pseudo-inverse. Many activation functions such as sigmoidal, sine, Gaussian or hard-limiting functions can be used at the hidden layer and the class is determined as the class which has the maximum output value. A comprehensive description of the S-ELM algorithm is given by Huang et. al., (2006).

Even though the ELM algorithm requires less training time, the random selection of input weights affects the generalization performance when the data is sparse or data is imbalanced. Suresh, Saraswathi and Sundararajan (2010) and Saraswathi et al. (2010) offer an improved version of ELM called the Sparse-ELM (S-ELM) which gives better generalization for sparse data. Hence, we use S-ELM algorithm for predicting the RSA of proteins where the imbalance in data varies with the different threshold values used. S-ELM is also well suited for RSA predictions of sequences whose structures have not yet been determined and where there are no homologs in existing sequences. The data is discus-

sed in detail in section 3.

We call the ELM algorithm for each of the training data sets over several thresholds. We find the optimal number of hidden neurons using a unipolar sigmoidal activation function (lambda = 0.001) and perform K-fold (k = 5) validations. In K-fold validation, the training set is separated into K-groups. K-1 groups are used for training in each of the K iterations and the model is tested on the remaining K^{th} group. The optimal parameters are stored and used during the testing phase. The performance of the S-ELM classifier and the time taken to develop the RSA S-ELM classifier model is compared with SVM using LIBSVM (Fan, Chen and Lin, 2005) approach to show that the S-ELM approach can achieve a slightly better performance within a much shorter time. Five-fold cross validation accuracies, processing time gains and comparative studies are discussed in the results section.

2.2 Data

Proteins consist of sequences of amino acid residues that play a key role in determining the secondary and tertiary structure of a protein. The sequential relationship among the solvent accessibilities of neighbouring residues can be used to improve the results (although solvent accessibility is considered evolutionarily less preserved than secondary structure). We use binary values and a window size of 8 to represent the amino acid sequences.

RSA of an amino acid residue is defined (Mucchielli-Giorgi et al. 1999) as the ratio of the solvent-accessible surface area of the residue observed in the 3-D structure to that observed in an extended tripeptide (Gly-X-Gly or Ala-X-Ala) conformation. RSA is a simple measure of the degree to which each residue in an amino acid sequence is exposed to its solvent environment. For our study, we consider the well-known Manesh data set (Manesh, Sadeghi, Arab, & Movahedi, 2001) which has a high imbalance with respect to the number of samples per class (Table 1), where the number of samples belonging to one class is much lesser than the samples belonging to the other classes.

The Manesh data set consists of 215 proteins, of which 30 proteins (7545 residues) with variable number of amino acid residues are used for classifier model development and the remaining 185 proteins (43137 residues) were used for evaluating the generalization performance of the S-ELM classifier through a 5-fold cross-validation model. The data in the training and testing set are cast into two-class and three-class problems (Table 1) by determining whether the RSA value is below, between or above a particular threshold. We use various % thresholds (0, 5, 10, 25, 50 for two-class and between 10_20 or 25_50 for three class), in order to compare our results with those existing in literature. A residue is considered as buried if its value is less than or equal to the lower range, partially buried if it is between the lower and the higher range and considered exposed if its RSA value is higher than the range of values (> 20 or > 50). The accuracy of the predictions depend on the value of the thresholds chosen and can vary widely with different residue compositions in different proteins as discussed in the results section.

Table 1: Samples per class for 2-class and 3-class data where thresholds are set between 0 and 50% for two class (C0 and C1) and between 0, 10 and 50 % for 3-class (C0 C1 and C2).

	Number of Training residues			Number of Testing residues		
%	C0	C1	C2	C0	C1	C2
0	867	6678	**	4713	38424	**
5	5796	1749	**	32943	10194	**
10	2826	4719	**	15864	27273	**
20	4065	3480	**	23111	20026	**
50	5796	1749	**	32945	10192	**
10_20	3888	831	2826	22265	5008	15864
25_50	1750	1750	4065	10194	9832	23111

3 RESULTS AND DISCUSSION

We compare the results of our simulation using S-ELM on the Manesh data set with the SVM algorithm and NETASA (Shandar & Gromiha 2002) methods (Figure 1 and Table 2), using the **same set** of proteins for training and testing. Hence comparisons with literature are made only with the NETASA results.

The accuracy of the RSA predictions is measured by the number of residues correctly classified as belonging to class1 (E for exposed) for the two class problem and as belonging to class2 (E) for the three class problem. Prediction accuracy for training and testing data sets is defined as the total number of correctly predicted values for each class over the total number of available residues in all classes. The data shown in Table 2 indicates that the S-ELM approach achieves a better accuracy for training and testing than the corresponding results for the

NETASA paper. The SVM algorithm takes a longer time to build the model as shown in Figure 2 and 3, whereas the S-ELM algorithm process data at the same speed for all combinations of data, showing that the algorithm does not slow down when complex data is involved. S-ELM uses optimal parameters that are stored during the training phase making it possible to run through the tests quickly.

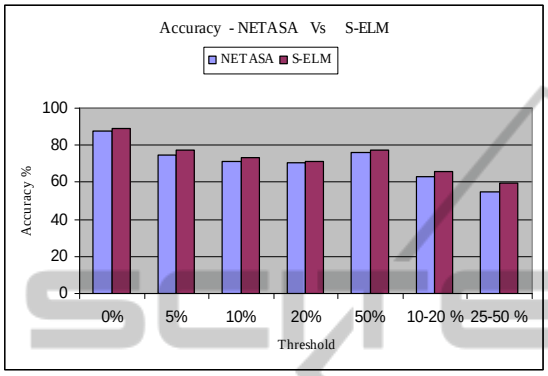

Figure 1: Accuracy comparison between NETASA and S-ELM, shows slight improvements for S-ELM method.

The training results for the SVM are between 89% and 99% for a range of thresholds. The corresponding testing results saw gains for some of the thresholds, while almost same results for the others. The test results vary from 69% to 89% over a range of thresholds, for the two class problem. The results are much better for the S-ELM algorithm, where the training and testing results are closer together showing better generalization. The training results vary between 73 % and 89% while the test results vary between 71% and 89% which are better than the results for the SVM and NETASA method. Our interest in including the SVM in our simulations was to show the advantages in time factor when the S-ELM algorithm is used. The training results for the S-ELM show a little gain over the NETASA and the SVM results, but the testing results for S-ELM clearly show higher results of between .006 to 4.476 % as seen in Figure 1 and Table 2. Similarly for the three-class problem, seen on the last two lines of Figure 3, the training accuracies for SVM are very high at 99% while the testing accuracies are 68% and 54% for two different thresholds, which are slightly higher than for the NETASA results.

For the S-ELM results, the training accuracies are closer to the testing accuracies, indicating better generalization for the 3-class problem also. Here the S-ELM test results show between 3 to 4% gains as compared to the NETASA results. As indicated by many results in the literature, the accuracies can vary widely for different thresholds and different number of classes into which the data is divided. A general trend in the literature is that the RSA prediction results vary between 70 % and 80%, similar to what is seen here. So, the S-ELM gives comparable results to literature.

Table 2: Training and Testing accuracies comparisons between NETASA, SVM and S_ELM for all thresholds using 350 hidden neurons are given. The support vectors are given for SVM data.

		NET-ASA	SVM	SVM	S-ELM
Thres - hold %		Accuracy %		SV	Accuracy for 350 hidden neurons %
Training	0	89.8	99.9	3837	88.6
	5	76.1	99.9	5894	79.8
	10	75.2	99.9	6610	74.0
	20	73.1	99.9	6826	72.98
	50	80.1	99.9	5897	79.80
	10 -20	65.1	99.9	7075	67.12
	25 -50	60.9	99.9	7087	63.51
Testing	0	87.9	89.1	**	**89.1**
	5	74.6	76.2	**	**77.3**
	10	71.2	71.2	**	**73.1**
	20	70.3	69.5	**	**71.3**
	50	75.9	76.3	**	**77.3**
	10-20	63	64.1	**	**66.0**
	25-50	55	58.1	**	**59.5**

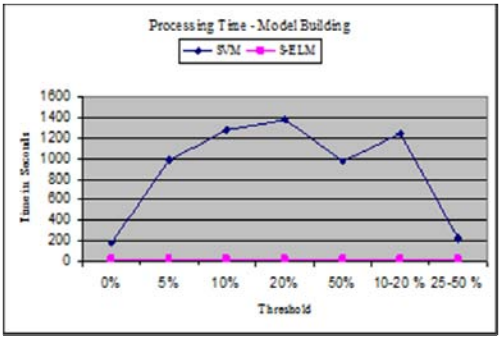

Figure 2: Processing time for modelling: SVM Vs. S-ELM, clear shows huge gains in time for S-ELM.

The biggest advantage of using S-ELM comes from the speed at which the data can be processed by the algorithm, while providing us with slightly better accuracies. It can be clearly seen from Table 3 that S-ELM has a clear advantage when it comes to processing speed. The same number of samples of

7545 training sample residues was used for model building for both algorithms. The ratio of time taken by SVM and S-ELM for model building, for the various thresholds range from 20.562 : 175 seconds which amounts to almost 8.51 times time gain by S-ELM for 0% threshold data. We find that the time gains range from 8 times to multiple folds, the highest being for the 20% threshold data where the ratio is 20.562:1372.2 which is a gain of over 66.734 times. Generally, the time taken for model building is most crucial, since the model needs to learn as much as possible in the shortest time.

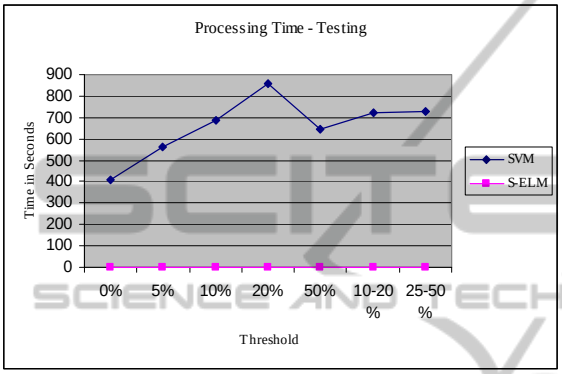

Figure 3: Processing time for testing: SVM Vs S-ELM.

For real time applications and for batch processing applications it might be useful to have faster testing capabilities and here we see that the S-ELM algorithm is much faster in its testing capabilities also. The same number of 43137 testing residues was used here for the test runs in both algorithms. Here the time gains between the testing times for 0 % threshold is .922:410 which amounts to 444.69 times fastr processing by S-ELM. We find similar gains for other thresholds with the highest gain for the 20% threshold at .937:857 which is 914.62 times faster processing speed. Both the SVM and the S-ELM were run on the same computer running XP windows operating system with 4 GB RAM and Matlab software.

Time taken for training and testing runs by SVM and S-ELM algorithms is given in Table 3. Figure 2 and Figure 3 illustrate the high processing time of SVM and the very low and steady processing times of S-ELM very clearly. The time taken by S-ELM is very low at less than one or two seconds, shown as a horizontal line close to the x-axis while the time taken by SVM is quite high, ranging between 200 and 1400 seconds for training and between 400 and 900 seconds for testing. S-ELM takes very little time for testing since stored optimal parameters are used to calculate the output analytically using ELM. There is no processing time data available to compare speeds with the NETASA method. Future studies will concentrate on increasing the accuracy of S-ELM further using optimization techniques to tune the S-ELM parameters for RSA prediction.

Table 3: Processing time for modelling, training and testing: comparison between SVM and ELM.

	SVM			S-ELM		
	Time in Seconds			Time in Seconds		
Threshold %	Modelling	Training	Testing	Modelling	Training	Testing
0	175	24.6	410	20.6	0.5	**0.92**
5	990	105	561	20.8	0.6	**0.94**
10	1273	67	686	20.9	0.6	**0.92**
20	1372	76	857	20.9	0.5	**0.94**
50	977	89	645	20.9	0.6	**0.95**
10-20	1239	88	723	**21**.0	**1.1**	**1.08**
25-50	226	74	728	**21**.0	**0.7**	**1.08**

4 CONCLUSIONS

We have used the SVM and S-ELM methods of classification for RSA prediction, using the Manesh data set. We have compared the performance of these algorithms with each other and with NETASA, with respect to the speed of processing and have shown that there are multiple-fold gains in computational efficiency while using S-ELM algorithm. It will be advantageous to use the S-ELM algorithm for real time and batch processing applications where accuracy and speed are equally important.

ACKNOWLEDGEMENTS

We acknowledge the support of National Institutes of Health through grants R01GM081680, R01GM072014, and R01GM073095 and the support of the NSF grant through IGERT-0504304.

REFERENCES

Adamczak, R., Porollo, A., & Meller, J. 2005. Combining prediction of secondary structure and solvent accessibility in proteins. *Proteins*, 59(3) 467-475.

Altschul, S., Madden, T., Schaffer, A., Zhang, J., Zhang, Z., Miller, W., & Lipman, D., 1997. Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. *Nucleic Acids Res*, 25(17) 3389-3402.

Berezin, C., Glaser, F., Rosenberg, J., Paz, I., Pupko, T., Fariselli, P., Casadio, R., & Ben-Tal, N., 2004. ConSeq: the identification of functionally and structurally important residues in protein sequences. *Bioinformatics*, 20, (8) 1322-1324.

Bondugula, R. & Xu, D., 2008. Combining Sequence and Structural Profiles for Protein Solvent Accessibility Prediction, *In Comput Syst Bioinformatics ConfC*, 195-202.

Carugo, O., 2003. Prediction of polypeptide fragments exposed to the solvent. *In Silico Biology*, 3(4), 417-428.

Chen, H., Zhou, H.-X., Hu, X., & Yoo, I., 2004 Classification Comparison of Prediction of Solvent Accessibility from Protein Sequences, *In 2nd Asia-Pacific Bioinformatics Conference (APBC)*, 333-338.

Cheng, J., Sweredoski, M., & Baldi, P., 2006. DOMpro: Protein Domain Prediction Using Profiles, Secondary Structure, Relative Solvent Accessibility, and Recursive Neural Networks. *Data Mining and Knowledge Discovery*, 13(1) 1-10

Cortes, C. & Vapnik, V., 1995. Support vector networks. *Machine Learning*, 20, 1-25.

David, M. P., Asprer, J. J., Ibana, J. S., Concepcion, G. P., & Padlan, E. A., 2007. A study of the structural correlates of affinity maturation: Antibody affinity as a function of chemical interactions, structural plasticity and stability. *Molecular Immunology*, 44 (6), 1342-1351.

Fan, R. E, Chen, P. H. and Lin, C. J., 2005. Working set selection using second order information for training SVM. *Journal of Machine Learning Research*, 6, 1889-1918.

Gianese, G., Bossa, F., & Pascarella, S., 2003. Improvement in prediction of solvent accessibility by probability profiles. *Protein Engineering Design and Selection*, 16(12) 987-992.

Huang, G. B., Zhu, Q. Y., & Siew, C. K., 2006. Extreme learning machine: Theory and applications. *Neurocomputing*, 70, (1-3) 489-501

Kim, H. & Park, H., 2004. Prediction of protein relative solvent accessibility with support vector machines and long-range interaction 3D local descriptor. *Proteins - Structure, Function, and Bioinformatics*, 54 (3), 557-562.

Manesh, N. H., Sadeghi, M., Arab, S., & Movahedi, A. A. M., 2001. Prediction of protein surface accessibility with information theory. *Proteins - Structure, Function, and Genetics*, 42 (4) 452-459.

Meshkin, A. & Ghafuri, H., 2010. Prediction of Relative Solvent Accessibility by Support Vector Regression and Best-First Method. *EXCLI*, 9, 29-38.

Mucchielli-Giorgi, M. H., Hazout, S., & Tuffery, P., 1999. PredAcc: prediction of solvent accessibility. *Bioinformatics*, 15 (2) 176-177.

Nguyen, M. N. & Rajapakse, J. C., 2005. Prediction of protein relative solvent accessibility with a two-stage SVM approach. *Proteins*, 59, (1) 30-37.

Ooi, T., Oobatake, M., Namethy, G., & Scheraga, H. A., 1987. Accessible surface areas as a measure of the thermodynamic parameters of hydration of peptides. *Proceedings of the National Academy of Sciences of the United States of America*, 84 (10) 3086-3090.

Petersen, B., Petersen, T. N., & Andersen , P., Nielsen, M., Lundegaard, C., 2009. A generic method for assignment of reliability scores applied to solvent accessibility predictions. *BMC Structural Biology*, 9:51.

Pollastri, G., Baldi, P., Fariselli, P., & Casadio, R., 2002. Prediction of coordination number and relative solvent accessibility in proteins. *Proteins*, 47(2) 142-153.

Pollastri, G., Martin, A., Mooney, C., & Vullo, A., 2007. Accurate prediction of protein secondary structure and solvent accessibility by consensus combiners of sequence and structure information. *BMC Bioinformatics*, 8, (1) 201

Saraswathi, S., Suresh, S., Sundararajan, N., Zimmerman, M. and Nilsen-Hamilton, M., 2010. ICGA-PSO-ELM approach for Accurate Cancer Classification Resulting in Reduced Gene Sets Involved in Cellular Interface with the Microenvironment. *IEEE Transactions in Bioinformatics and Computational Biology*, http://www.computer.org/portal/web/csdl/doi/10.1109/TCBB.2010.13.

Suresh, S., Saraswathi, S., Sundararajan, N., 2010. Performance Enhancement of Extreme Learning Machine for Multi-category Sparse Data Classification Problems. *Engineering Applications of Artificial Intelligence*, http://dx.doi.org/10.1016/j.engappai.2010.06.009.

Shandar, A. & Gromiha, M. M., 2002. NETASA: neural network based prediction of solvent accessibility. *Bioinformatics*, 18(6), 819-824.

Shen, B. & Vihinen, M., 2003. RankViaContact: ranking and visualization of amino acid contacts. *Bioinformatics*, 19(16), 2161-2162.

Sim, J., Kim, S.-Y., & Lee, J., 2005. Prediction of protein solvent accessibility using fuzzy k -nearest neighbor method. *Bioinformatics*, 21(12), 2844-2849.

Singh, Y. H., Gromiha, M. M., Sarai, A., & Ahmad, S., 2006. Atom-wise statistics and prediction of solvent accessibility in proteins. *Biophysical Chemistry*, 124(2), 145-154.

Wagner, M., Adamczak, R., Porollo, A., & Meller, J., 2005. Linear regression models for solvent accessibility prediction in proteins. *J Comput Biol*, 12(3), 355-369.

Wang, J. Y., Lee, H. M., & Ahmad, S., 2007. SVM-Cabins: prediction of solvent accessibility using accumulation cutoff set and support vector machine. *Proteins*, 68(1), 82-91.

Zarei, R., Arab, S., & Sadeghi, M., 2007. A method for protein accessibility prediction based on residue types and conformational states. *Computational Biology and Chemistry*, 31(5-6) 384-388.